

# Lecture 7&8 -2

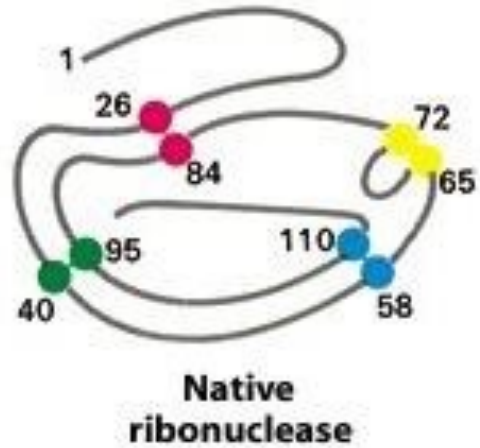
**Bio312**

Instructor: Dr. Lanlan Han

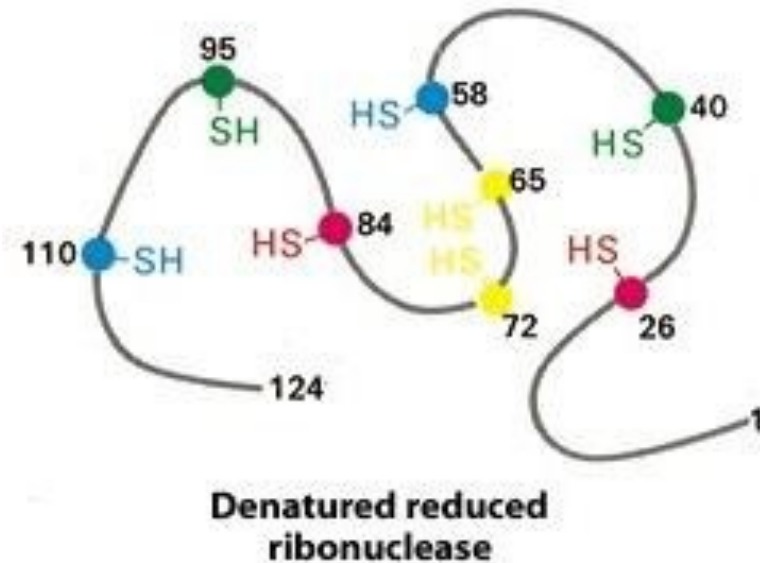
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# Anfinsen Experiment: Review

- The primary structure determines its tertiary structure.

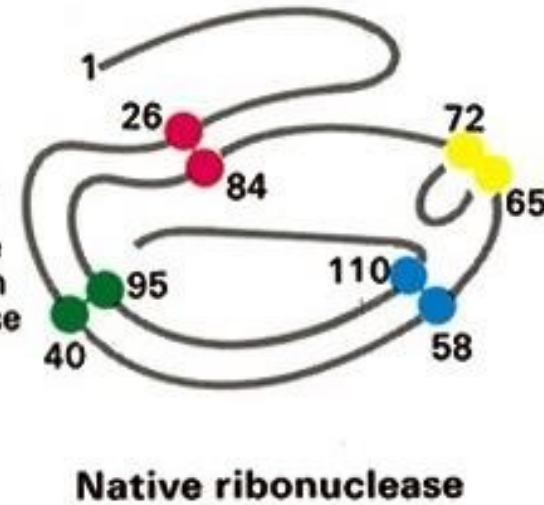


8 M urea and  $\beta$ -mercaptoethanol

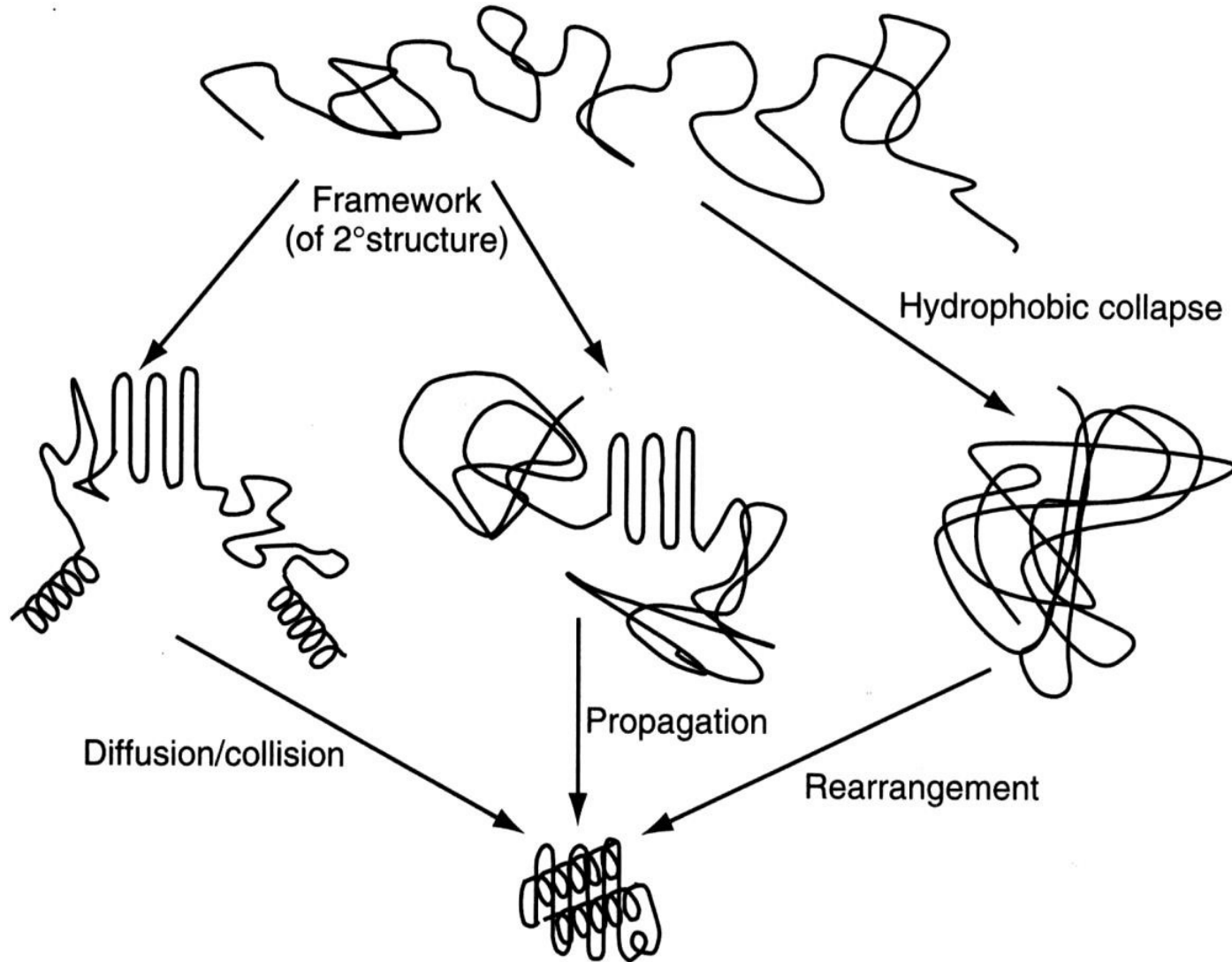


Dialysis to remove urea and  $\beta$ -mercaptoethanol

Air oxidation of the sulfhydryl groups in reduced ribonuclease

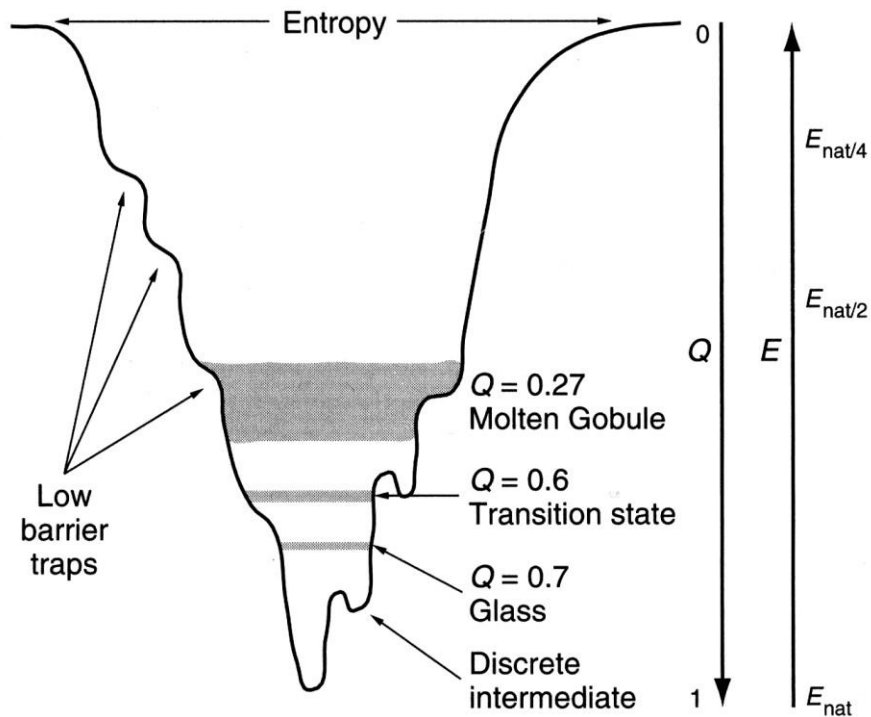


# Three Classic Models of Protein Folding: Review

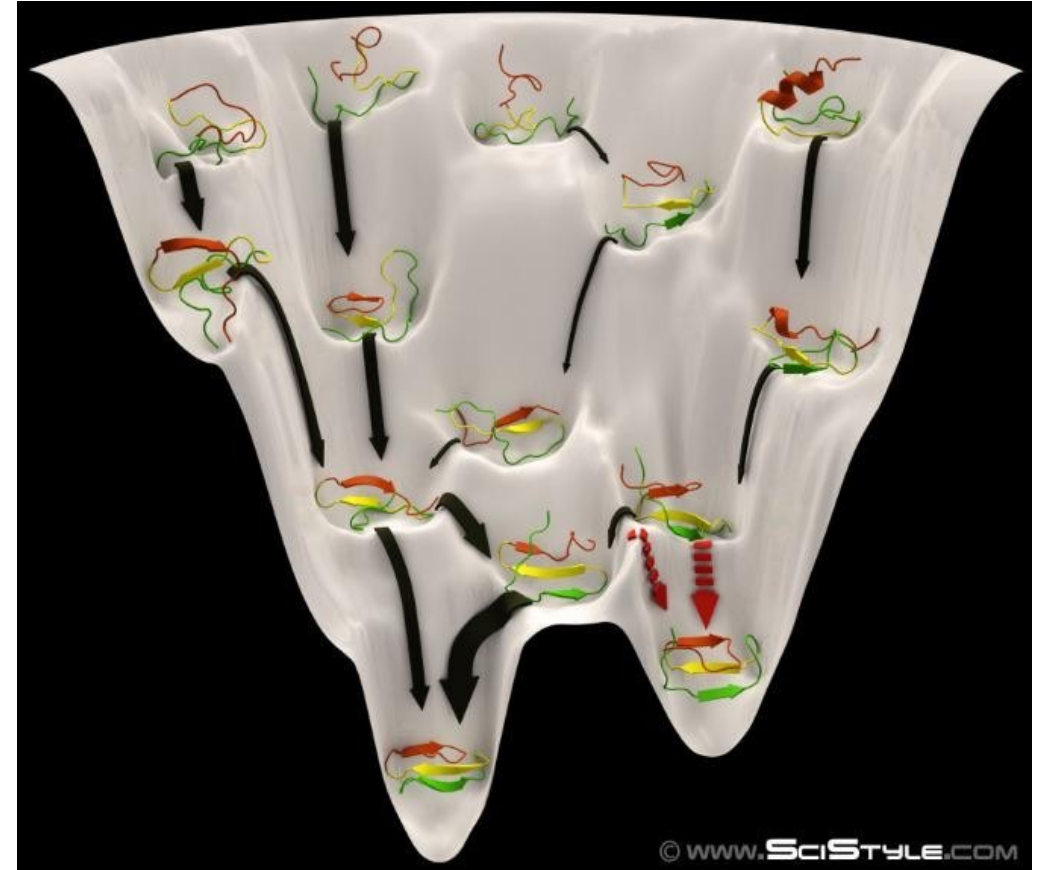


# The Folding Funnel: Review

- The funnel is a collection of **geometrically similar structures** to native proteins.



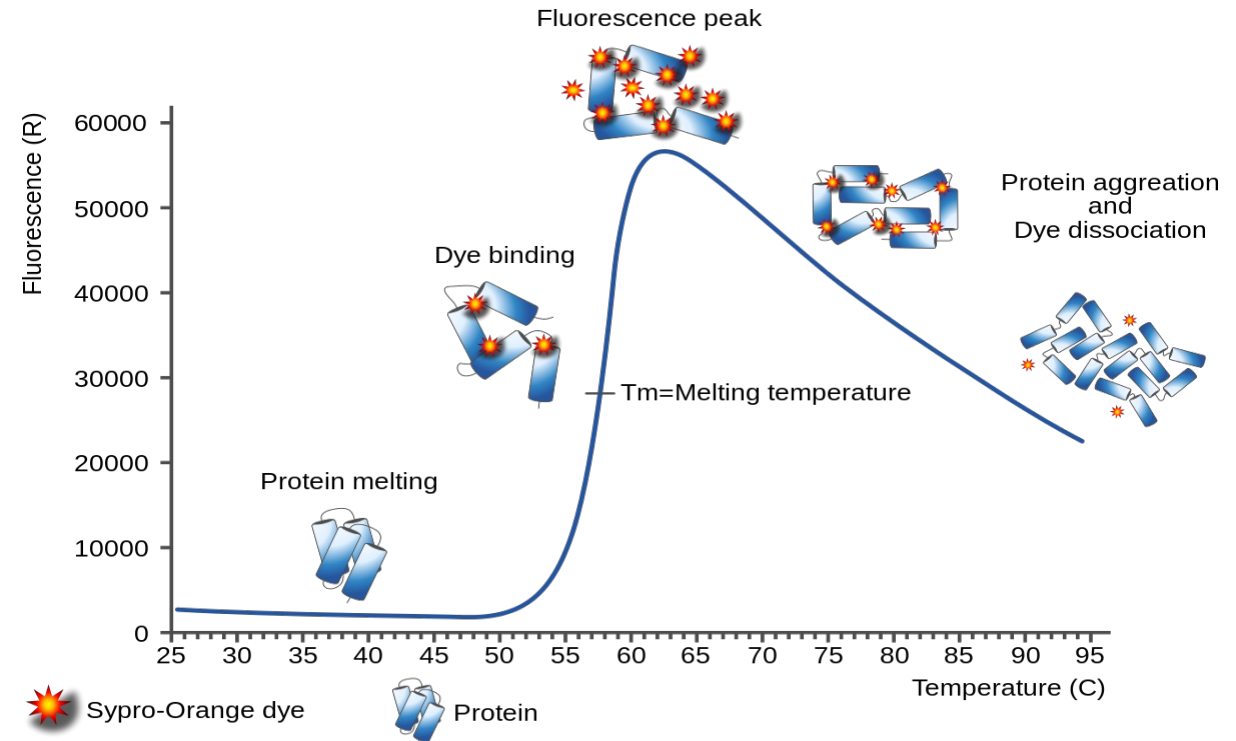
**Figure 19.16** Cross section through a folding funnel.  $E$  corresponds to free energy.  
[Courtesy of P. G. Wolynes]



Protein's native state is its **free energy minimum**

# Thermal Shift Assay: Review

- *Principle:* Protein hydrophobic core is buried. When the protein solution is heated up, the protein is denatured, and hydrophobic region is exposed to SYPRO™ orange fluorescent dye. The fluorescent signal can be determined in real time instrument.



# Circular Dichroism (CD) for Protein Structure: Review

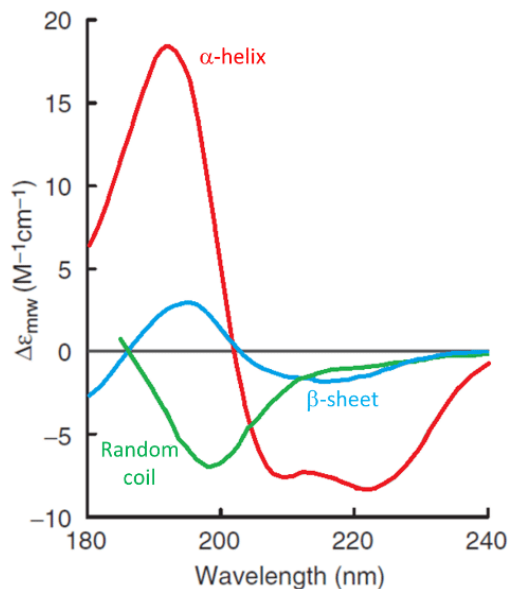
- Different wavelength probe different levels of structure:

1. **Far UV** (smaller wavelengths, 190-250 nm)

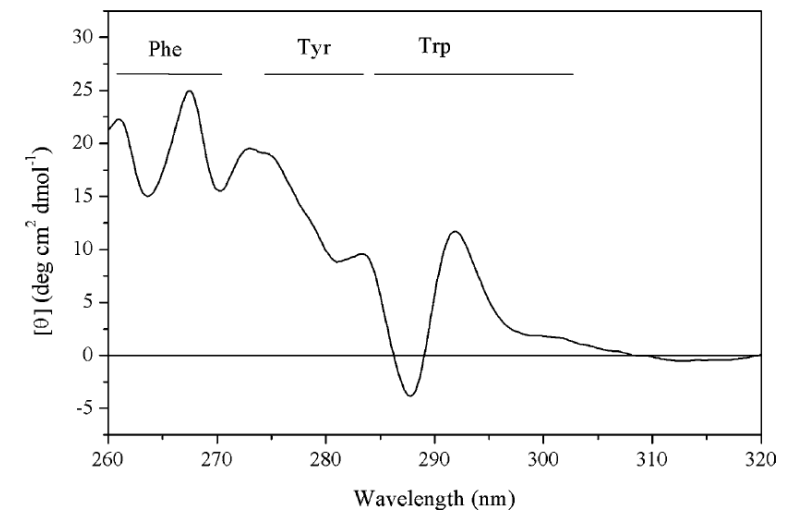
- probing different **secondary structures** in a protein

2. **Near UV** (longer wavelengths, 260-320 nm)

- Probing aromatic amino acids--> **tertiary structure change information**

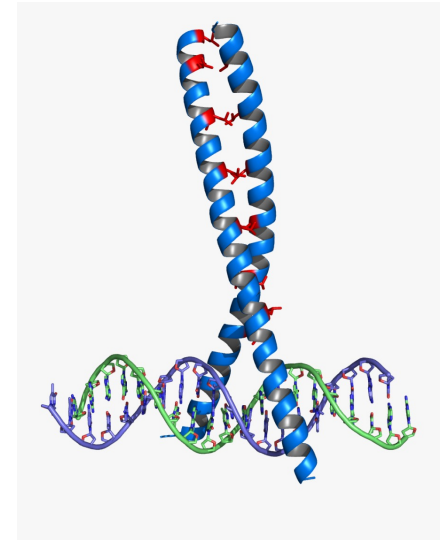
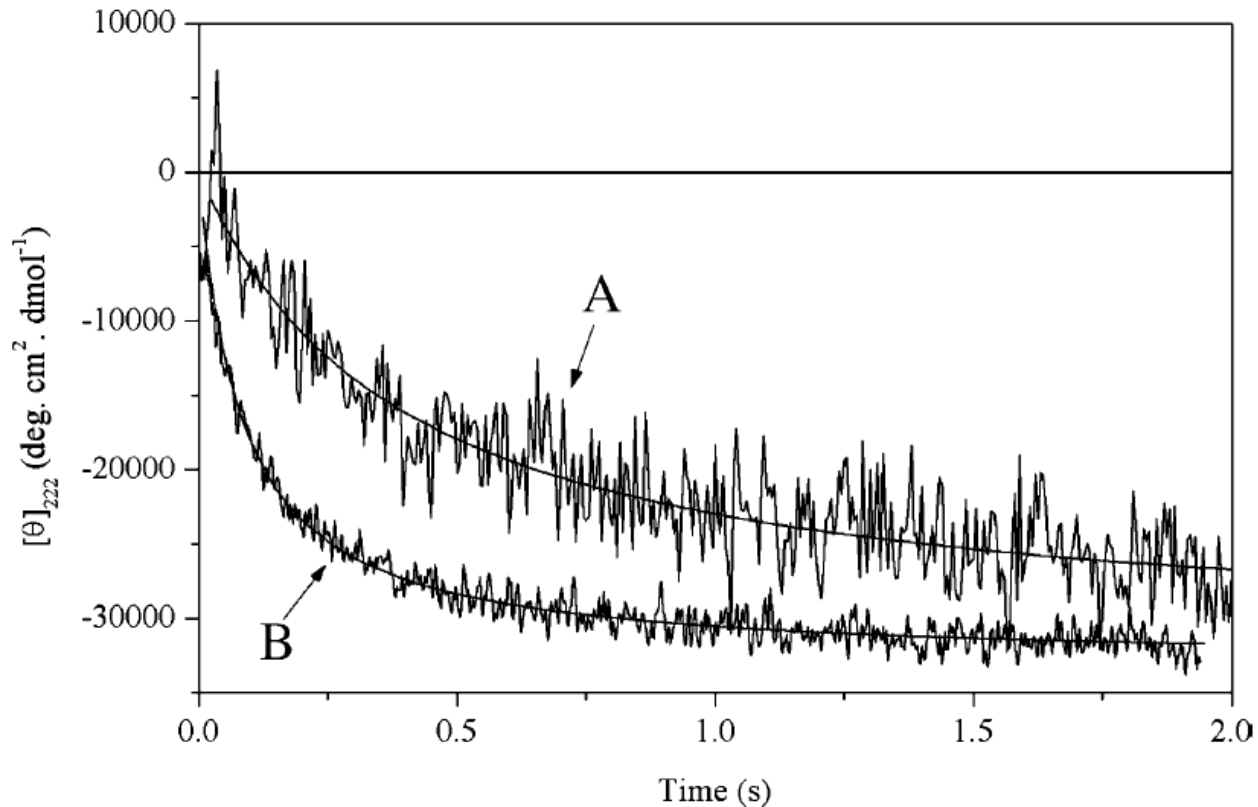


- **$\alpha$  helix** : negative bands at 208 and 222 nm and positive band at 193 nm
- **$\beta$  sheet** : negative band at 218 nm and positive band at 195 nm
- **Random coil**: negative band at 195 nm and low ellipticity above 210 nm



# 4. Structural Stability (Folding or Unfolding)

The refolding of a leucine zipper peptide monitored by stopped flow CD at 222 nm.

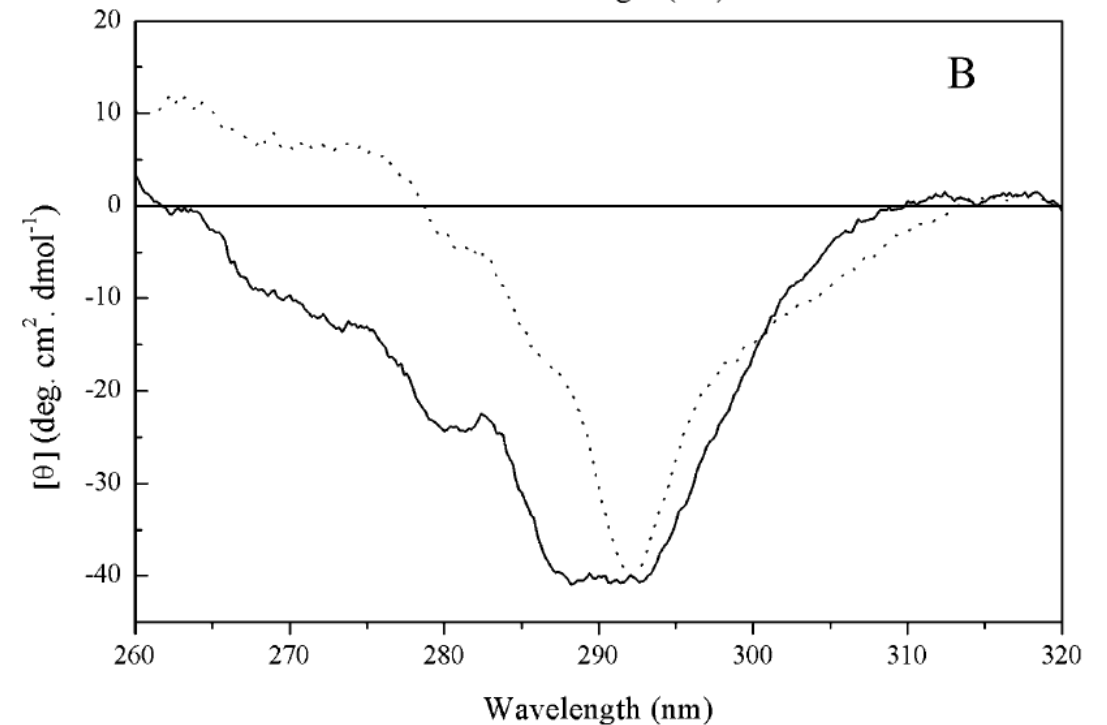
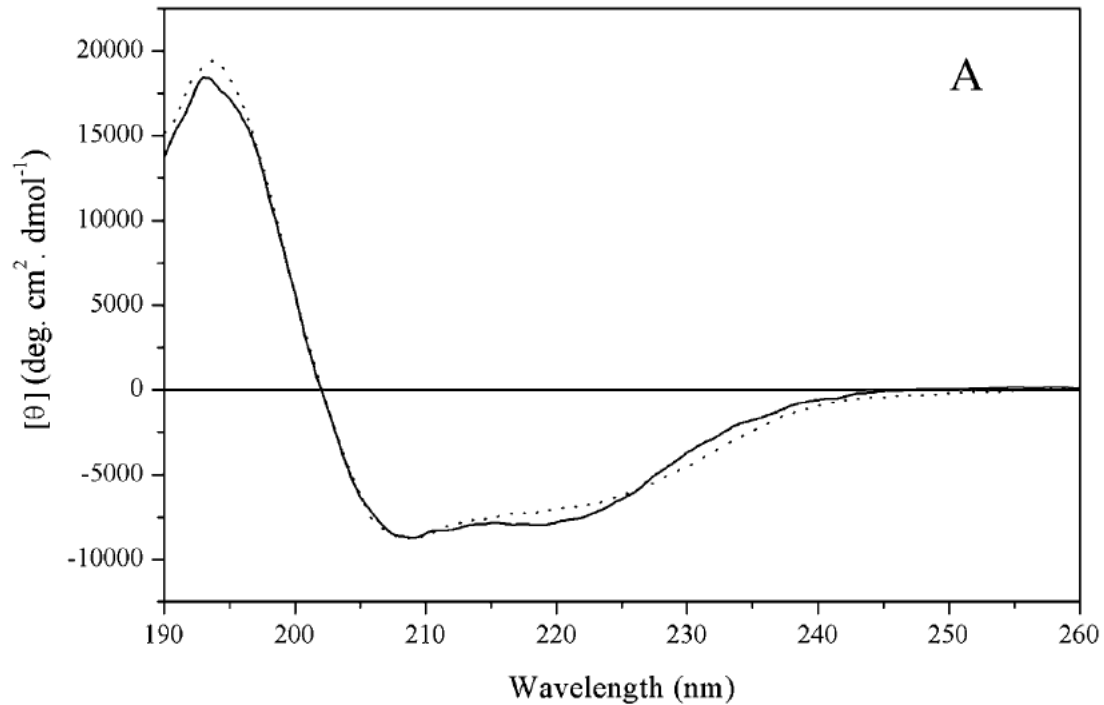


33-residue peptide at concentrations of 6  $\mu\text{M}$  (A) and 26  $\mu\text{M}$  (B) was refolded after denaturation in GdmCl.

# 5. Conformational Changes in Protein-Ligand Binding<sub>1</sub>

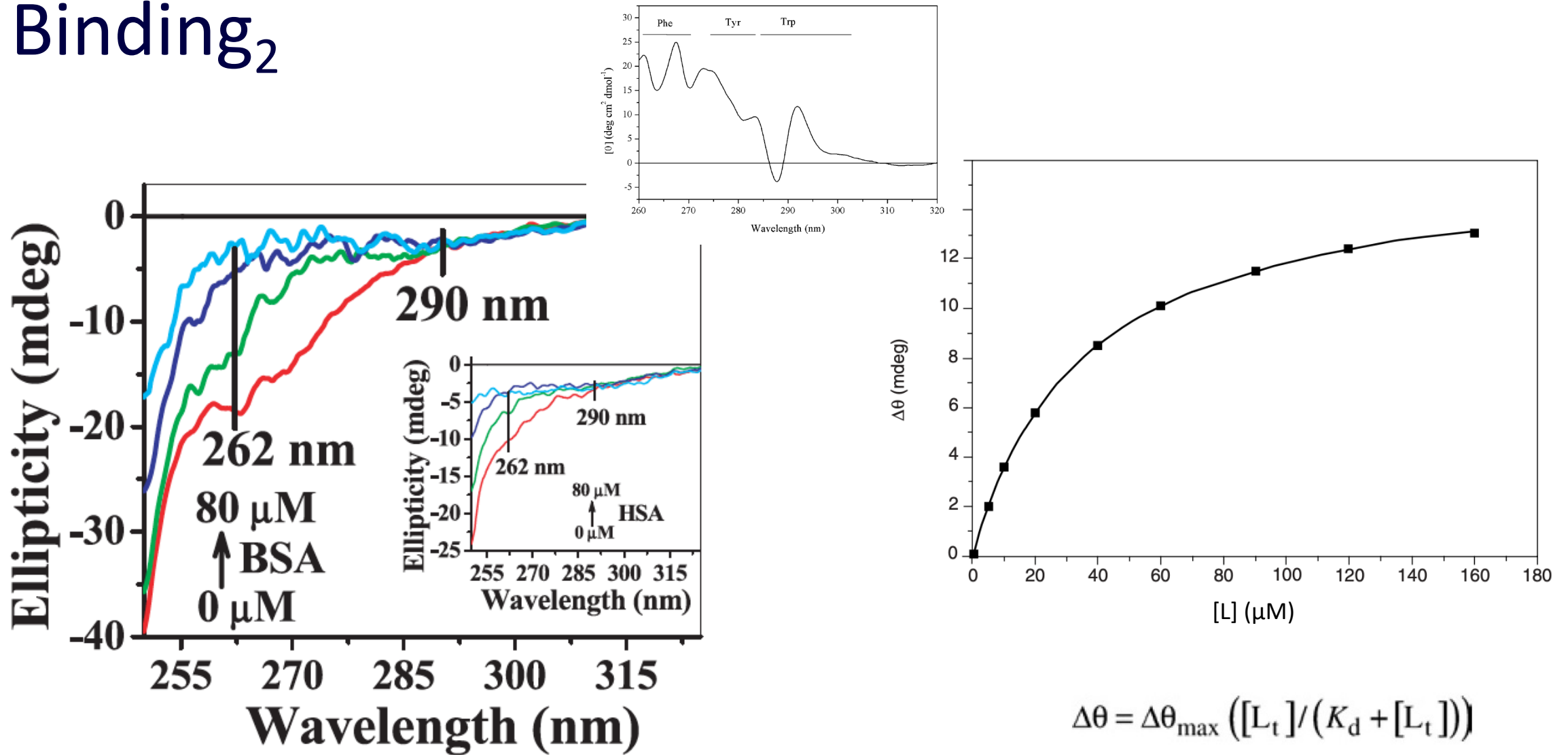
molybdate-sensing protein ModE (solid line)

molybdate-sensing protein ModE + 1 mM molybdate (dotted line)





# 5. Conformational Changes in Protein-Ligand Binding<sub>2</sub>

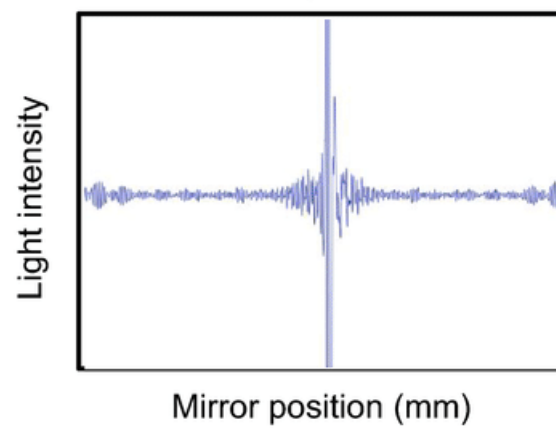
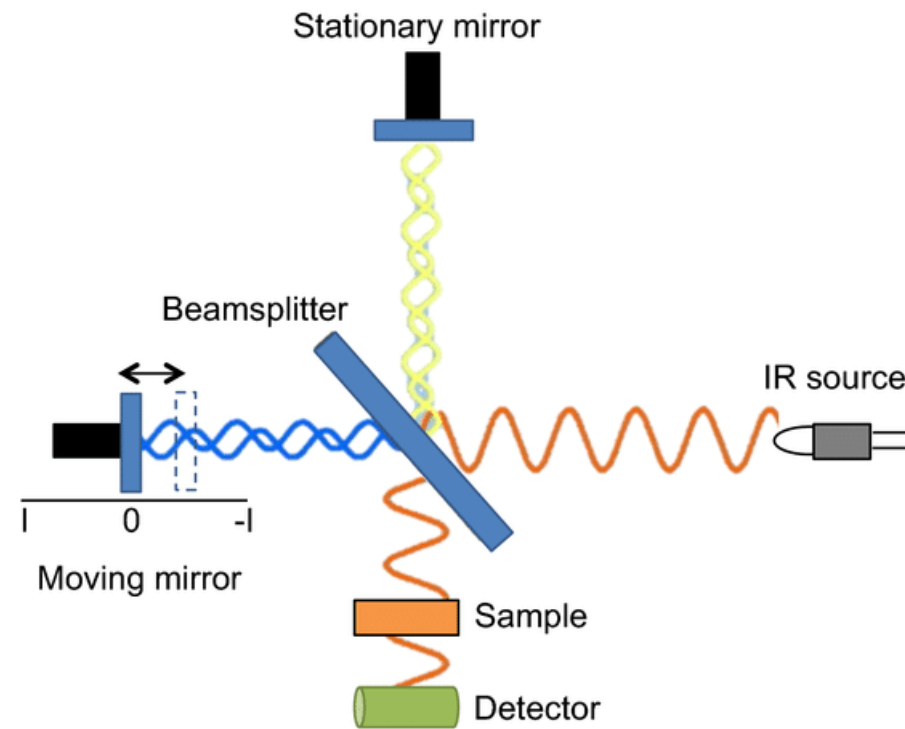


$$\Delta\theta = \Delta\theta_{\max} \left( \frac{[L_t]}{K_d + [L_t]} \right)$$

# Infrared (IR) Spectroscopy

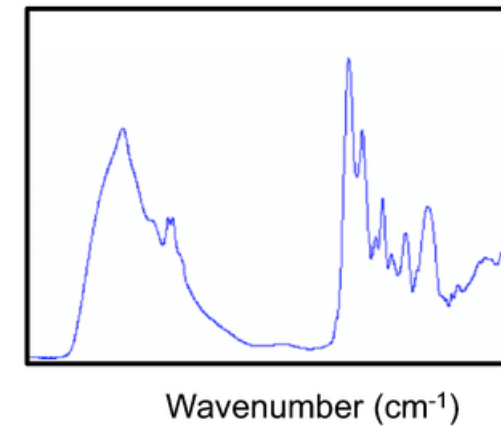
- IR spectroscopy studies the interaction between matter and infrared radiation.
- IR radiation is absorbed at characteristic wavelengths (wave number,  $1/\lambda$ ) by different groups.
  - C=O groups absorb in the IR at 1550-1750  $\text{cm}^{-1}$
- IR is not a great technique for studying proteins, because water absorbs IR radiation strongly in regions that overlap the protein absorbance bands.

# Fourier Transform IR (FTIR)



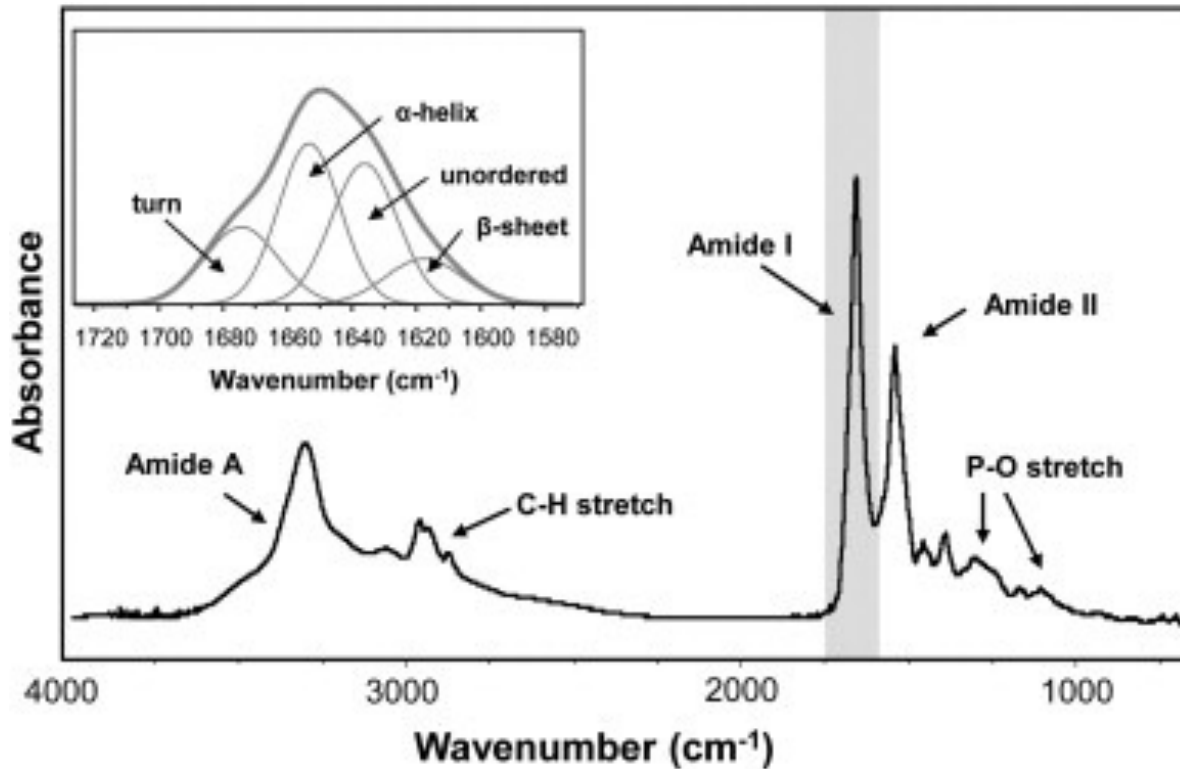
**INTERFEROGRAM**

FFT



**IR SPECTRUM**

# FTIR Spectra



- FTIR can be used to determine the amounts of each kind of 2<sup>o</sup> structure and the conformation analysis.
- Amide I and amide II band shape is very characteristic for a protein's 2<sup>o</sup> structure.

# Discussion

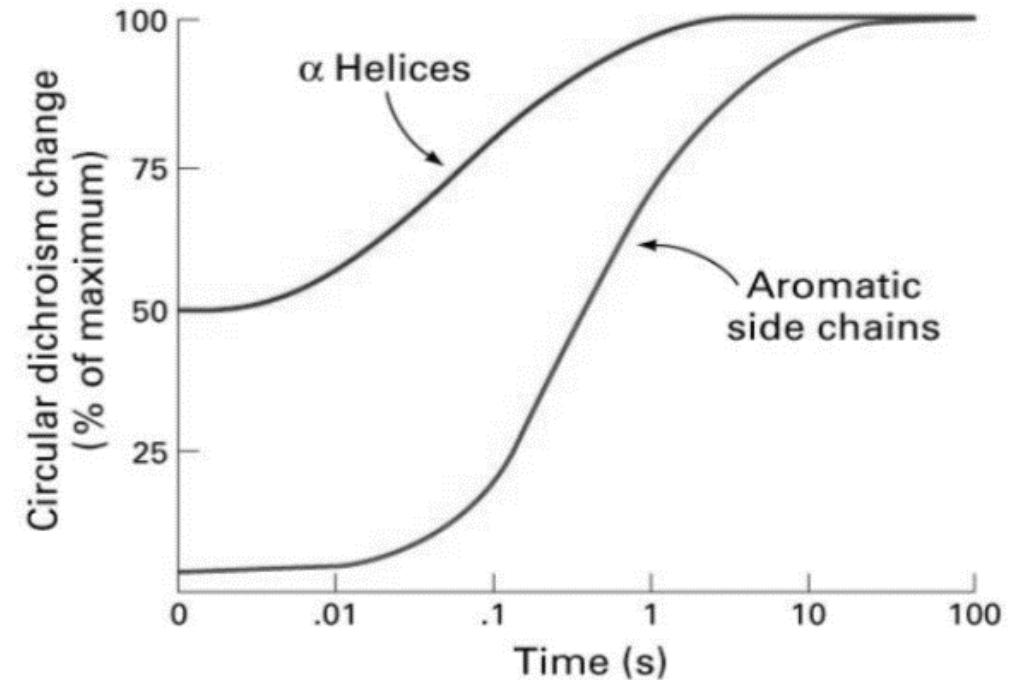
The figure on the right is the folding profile of protein X as monitored by circular dichroism. Among the three classic models of protein folding, which refolding model is:

(a) *least* suited for protein X? Explain your reasons.

hydrophobic-collapse Model

(b) *best* suited for protein X? Explain your reasons.

Nucleation Model



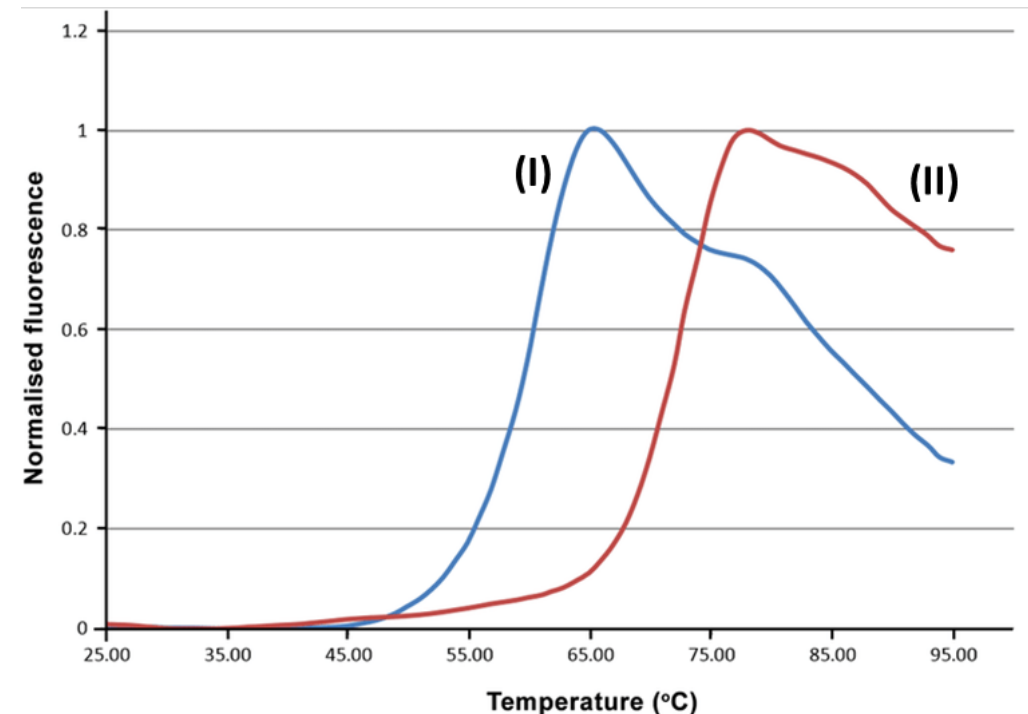
# Practice

A thermal shift assay on an enzyme “Y” in the *absence and presence* of 1 mM of Inhibitor A, which has a known  $K_i$  value of 1  $\mu\text{M}$  was performed. The result of the assay is shown in the figure.

Based on your knowledge on the thermal shift technique, which denaturation curve ((I) or (II)) in the figure is more likely to be the denaturation curve of:

(a) the apo-enzyme? 1

(b) enzyme Y + 1 mM Inhibitor A? 2

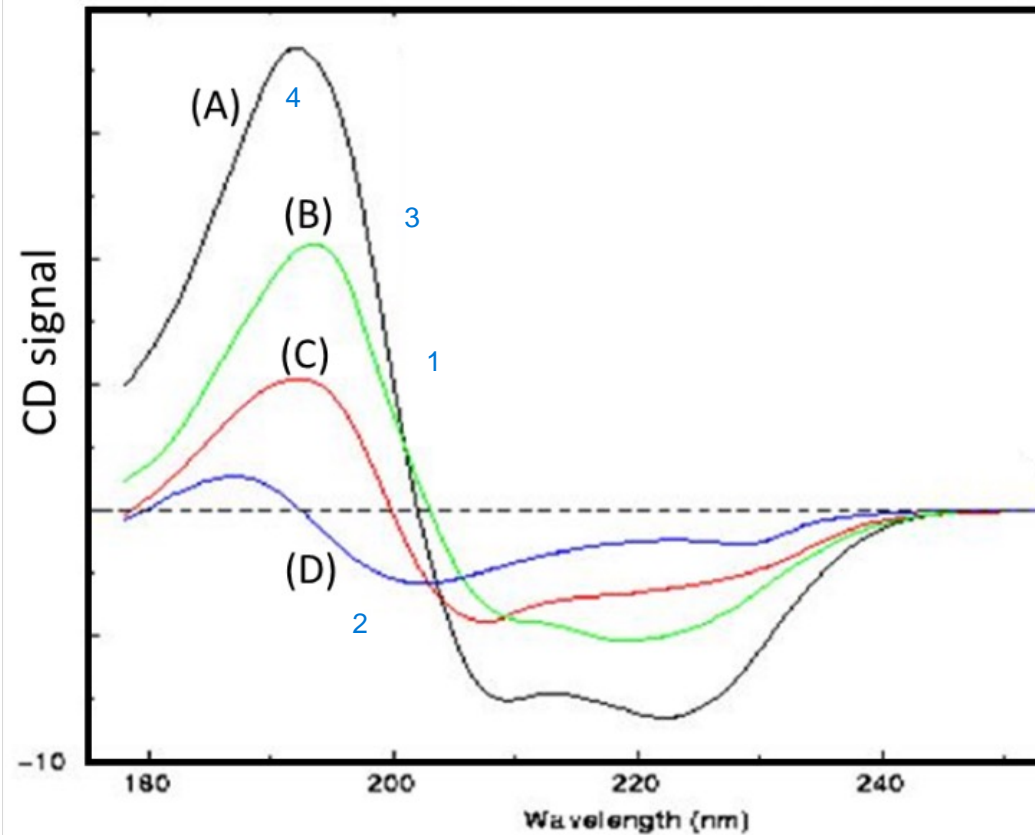


# Practice

Which spectrum (A, B, C or D) is most likely to belong to:

lysozyme? chymotrypsin?

myoglobin?



1. Lysozyme (40% helix, 20%  $\beta$  sheet)
2. Chymotrypsin (10% helix, 25%  $\beta$  sheet)
3. Triosephosphate isomerase (75% helix, 15%  $\beta$  sheet)
4. Myoglobin (96% helix)

